

Microbiology Collection & Transport Guide

Overview

Focus Diagnostics offers routine and special cultures for bacteria, viruses, mycobacteria and fungi. (Please refer to pages 9-11 for detailed information on Virology Collection and Transport procedures.) The Microbiology Department utilizes a variety of methods for identification including, but not limited to, traditional biochemicals and enzyme-based methods, serotyping, genetic probes, microscopy and High Performance Liquid Chromatography (HPLC). Antimicrobial susceptibility testing is available for bacteria, fungi, Herpes Simplex Virus, and mycobacteria. Antimicrobial, antifungal, antiviral, and antiretroviral drug level testing is performed with bioassay, HPLC, and spectrophotometric methods. It is the goal of the Microbiology Department to utilize the most current methods available to provide you with an accurate and rapid result.

Test Request Information

Focus' Microbiology Department requests that as much information as possible be provided. This should include source, date collected, antibiotics that the patient is currently taking, and organism identification when appropriate.

Collection, Transportation and Storage Guidelines

Specimens should be collected in the acute phase of infection. Transport all specimens to the laboratory as soon as possible, usually within 24 hours post collection. The charts included in this section describe specimen collection and optimum transport procedures by body site and by organism. This information serves as a guide only. Please do not hesitate to contact Focus' Microbiology Department for further details regarding the collection and transport of specimens.

Reporting

The availability of preliminary and final reports varies according to the type of culture.



Microbiology Specimen Collection & Transport by Organism

(See Pages 6 to 11 for Mycology, AFB, and Virology Specimens)

Organism	Test	Specimen Source	Optimum Transport Procedure <i>(Transport all specimens within 24 hours or overnight unless otherwise specified.)</i>
Acanthamoeba	Acanthamoeba Culture	Corneal scrapings, conjunctival scrapings, contact lens and contact lens fluid (recommended)	Place scrapings or tissue in sterile saline. Room Temperature
Anaerobes	Culture, Anaerobe	Deep wounds, sterile fluids, abscess material, trans-tracheal aspirates, and tissue	Anaerobic transport Room Temperature
Bartonella	Bacterial Culture, Special	Whole blood (EDTA), tissue, CSF, lymph node aspirate, or joint fluid	Blood preferred; tissue in BHI or Thioglycollate. Room Temperature
Bordetella pertussis/ parapertussis	Bordetella pertussis/ parapertussis Culture	NP swab is the preferred specimen. Other acceptable sources are bronchial or NP secretions or aspirates and transtracheal aspirates on appropriate transport swab. Use calcium alginate or Dacron-tipped NP swabs only.	Charcoal transport swab Room Temperature <i>Note: A PCR is strongly recommended if the specimen is not received <24h post collection. Alternatively, a DFA may be run but is less sensitive.</i>
Borrelia spp.	Borrelia burgdorferi (Lyme) Culture Borrelia hermsii Culture	Blood (preferred), skin biopsy at lesion periphery, CSF, joint fluid or live tick	Blood preferred. Room Temperature
Brucella	Bacterial Culture, Special	Blood, bone marrow, lymph node aspirates, synovial fluid, CSF, abscess aspirates, and liver or spleen biopsy	Room Temperature <i>Note: anticoagulants may affect organism viability upon prolonged storage.</i>
Campylobacter	Bacterial Culture, Aerobic (Stool)	Stool, rectal swab	Cary-Blair transport medium 2-8° C
E. coli	E. coli, Enterohemorrhagic E. coli, Enteroinvasive E coli, Enteropathogenic E. coli, Enterotoxigenic	Pure isolate	Room Temperature
Francisella tularensis	Bacterial Culture, Special	Lymph node aspirate, sputum, throat swab, bronchial washing, lesion biopsy	FREEZE , or Amies gel swab at Room Temperature.
Haemophilous ducryei	Bacterial Culture, Special	Genital lesion	Amies gel swab preferred 2-8° C

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Organism	Test	Specimen Source	Optimum Transport Procedure <i>(Transport all specimens within 24 hours or overnight unless otherwise specified.)</i>
Helicobacter pylori	H. pylori Culture	Gastric biopsy	20% glycerol medium (preferred), or Stuart's or Cary Blair transport medium. 2-8° C
Legionella spp.	Legionella Culture	Sputum, transtracheal aspirate, bronchoscopy specimen, biopsy, CSF, pericardial fluid, pleural fluid, peritoneal fluid	Sterile container 2-8° C
Leptospira spp.	Leptospira Culture	Blood (heparin, oxalate or citrate), CSF (first week of illness), urine (after first week of illness).	Sterile container. Neutralize urine to pH of 7.0 by adding sterile dilute acid or base. 5-20° C
Listeria monocytogenes	Bacterial Culture, Special	Blood, CSF, amniotic fluid, placenta or fetal tissue	2-8° C
Mycoplasma pneumoniae	Mycoplasma pneumoniae Culture	Throat or nasopharyngeal swab/wash, sputum, tracheal aspirate	Mycoplasma transport medium 2-8° C
Neisseria gonorrhoeae	Bacterial Culture, Special	Genital tract, urine, anal area, oropharynx, conjunctiva, Bartholin's gland, fallopian tubes, endometrium, blood, joint fluid, skin lesions, or gastric contents of neonates.	N. gonorrhoeae transport system Room Temperature
Streptobacillus moniliformis	Bacterial Culture, Special	Blood or joint fluid	Blood in non-SPS medium, ASAP, Room Temperature
Ureaplasma urealyticum/ Mycoplasma hominis	Mycoplasma Culture, Genital	Urethral, vaginal or cervical swab, urine, abscess, prostatic secretions, placenta, fetal tissue from spontaneous abortions or still births, endometrial washings or tissue biopsy, fallopian tube tissue, respiratory specimens	Mycoplasma transport medium 2-8° C
Vibrio	Bacterial Culture, Special	Stool prior to antibiotic therapy	Cary-Blair transport medium 2-8° C
Yersinia pestis	Bacterial Culture, Special	Lymph node aspirate, sputum, blood, throat swab/washings, CSF	2-8° C

*Please note that this is a suggested specimen collection guide.
For specimen types that are not listed, please contact Focus' Scientific Director of Microbiology.*